

Review

Anti-Oxidative Therapy in Diabetic Nephropathy

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Abstract

Chronic kidney disease is generally progressive and currently has no reliable treatment to reverse a decline in kidney function or to slow the progression of the disease. Diabetic nephropathy is one of the leading causes of end-stage kidney failure. Kidney damage in diabetic nephropathy is largely attributed to the increased oxidative stress, affecting its metabolic activity, metabolic pathways, and hemodynamic pathways. In diabetic patients, hyperglycemia causes an increase in the production of reactive oxygen species that further increase oxidative stress. These reactive oxygen species are created through a variety of pathways, providing the opportunity for treatment using anti-oxidative defense mechanisms to prevent vascular injury. This review will give an overview of oxidative stress, along with the current treatments and limitations of diabetic nephropathy. We will also discuss the potential of antioxidative therapies, with an emphasis on the nuclear factor erythroid 2-related factor 2 (Nrf2) pathway.

Keywords: review; diabetic nephropathy; antioxidative therapies; oxidative stress; diabetes; hyperglycemia; hypertension

1. Introduction

Diabetic nephropathy (DN) is one of the serious complications of diabetes occurring in 20% to 40% of diabetic patients and is one of the most common causes of end-stage renal disease (ESRD) [1]. Currently, there is no curative therapy for DN. Dialysis and kidney transplantation are typically required for advanced chronic kidney disease (CKD) or ESRD.

Evidence suggests that the progression of DN is a result of metabolic and hemodynamic interactions and dysregulation [2]. As such, DN treatment plans typically include glycemic and hypertensive care to help reestablish metabolic and hemodynamic control to delay renal injury. However, there is no single hypertensive therapy that is a “one size fits all” for every patient, and aggressively pursuing a lower blood pressure in patients with pre-existing cardiovascular diseases is often contraindicated [3]. Additionally, tight control of blood pressure may contribute to the slower progression of DN to ESRD but cannot completely halt it.

Given the lack of definitive treatment for DN, there is a need for new therapeutic agents and targets that can either reduce the risk, further slow, or outright prevent the progression of DN to full-blown ESRD. One such approach aims to mitigate the deleterious effects of DN-associated oxidative stress on the kidney by increasing renal cell antioxidant capacity.

Oxidative stress has been shown to play a central role in the pathogenesis of DN [4]. The metabolic and hemodynamic dysregulation that are characteristic of diabetes result

in the accumulation of intracellular reactive oxygen species (ROS) leading to impaired cellular function and chronic disease [4–6]. Thus, a therapeutic approach aimed at increasing the antioxidative capacity of renal cells under diabetogenic conditions is a promising approach in the treatment of DN. Nuclear factor erythroid 2-related factor 2 (Nrf2), the central regulator of the cellular response to oxidative stress, is a promising target for therapies aimed at increasing renal cell antioxidant capacity in patients with DN [7].

In this review, we discuss the role of oxidative stress in the pathogenesis of DN, the Nrf2 pathway, and pre-clinical antioxidative therapies that have been shown to slow the progression of diabetic kidney disease [4,8,9].

2. Diabetic Nephropathy and Oxidative Stress

2.1 Renal Impairment

DN is the main cause of ESRD and it is identified based on renal histological and functional alterations [10]. Diabetic insults induce microvascular injuries within the glomeruli and tubulointerstitial compartment [11]. In the glomeruli, thickening of basement membranes, mesangial expansion, hypertrophy, and loss of podocytes, specialized epithelial cells, occurs alongside the expansion of the tubular basement membranes, tubular atrophy, interstitial fibrosis, and arteriosclerosis [11].

Hyperglycemia and hypertension are two major insults that result in the decline of renal function, and both occur through different cellular pathways [12,13]. Hyperglycemia causes renal impairment through inducing renal



Table 1. List of reactive oxygen species accompanied by their formula.

ROS	Formula
superoxide anion	O ₂ ⁻
hydrogen peroxide	H ₂ O ₂
hydroxyl radical	•OH
singlet oxygen	O=O
peroxyl radical	ROO•
alkoxyl radical	R-O•
lipid hydroperoxide	LOOHs
peroxynitrite	ONOO ⁻
hypochlorous acid	HOCl
nitric oxide	NO

R indicates any carbon groups, L indicates any lipids, and NO is grouped as a reactive nitrogen species that is relevant to diabetic nephropathy.

hypoxia, oxidative stress, altering renal vasoreactivity, and volume depletion due to osmotic diuresis [14]. Systemic arterial hypertension affects the glomerulus through altering hydraulic pressure which has been associated with relative arteriolar dilation and efferent arteriolar constriction [15,16]. Hyperlipidemia is also considered to play a significant role in renal damage that leads to DN [17]. While DN is a result of a combination of several factors, downstream effects of hyperglycemia, hypertension, and hyperlipidemia converge at oxidative stress, which has been shown to play a central role in the progression of DN; thus, targeting oxidative stress may be an important approach for the treatment of DN [18].

2.2 Oxidative Stress

ROS are molecules containing oxygen, many of which are free radicals, such as superoxide, hydrogen peroxide, hydroxyl radical, singlet oxygen, peroxyl radical, alkoxyl radical, lipid hydroperoxide, peroxynitrite, hypochlorous acid, ozone, and nitric oxide (Table 1) [19]. Due to the kidney's high metabolic activity, various types of cells including endothelial, vascular smooth muscle, mesangial, and tubular epithelial cells are able to produce ROS [20,21]. Intrarenal oxidative stress is associated with the initial stages and progression of DN [22].

2.3 The Role of Hyperglycemia and Oxidative Stress in DN

The production of ROS is increased during hyperglycemia, which further worsens diabetic complications [23,24]. Oxidative stress from hyperglycemia causes metabolic modifications of the target tissue and changes in renal hemodynamics that increases vascular injury [11,25]. Thus, oxidative stress plays a significant role in the pathogenesis and progression of DN [26]. ROS are a major contributor to diabetic vascular damage and the overproduction of these species is directly linked to hyperglycemia [26].

In hyperglycemic conditions, the overproduction of ROS can be induced through different mechanisms based on cytosolic and mitochondrial sources [11]. Mitochondrial sources of ROS are oxidative phosphorylation, uncoupling of the respiratory chain, and dysregulation of complex I, coenzyme Q, and complex III [11,27–30]. The metabolism of glucose produces energy for the mitochondrial respiratory chain via oxidative phosphorylation by converting it to pyruvate in order to be reduced into NADH and FADH₂. NADH is responsible for donating electrons to the respiratory chain and in hyperglycemic conditions, the NADH/NAD⁺ ratio is increased in stressed porcine endothelial cells [31]. It is important to note that free fatty acids can produce NADH and FADH₂ via the tricarboxylic acid cycle, thus excess free fatty acids can mimic hyperglycemic damage to the mitochondria [11]. *In vitro* studies have shown that ROS production has been prevented by inhibiting the electron transport chain complex II via uncoupling oxidative phosphorylation, protein-1, and manganese superoxide dismutase [32]. It was shown that controlling mitochondrial ROS production the formation of advanced glycation end-products (AGE) was also inhibited [32]. AGEs are capable of inducing apoptosis and overexpressing vascular endothelial growth factor (VEGF) and monocyte chemoattractant protein-1 (MCP-1) [33]. VEGF and MCP-1 are linked to early phases of DN [33]. AGEs have been shown to increase ROS production and inhibit anti-oxidative protection mechanisms but there is also evidence that ROS can cause the production of some AGEs [34].

Cytosolic sources can be attributed to non-enzymatic pathways such as advanced glycation and glucose auto-oxidation or through enzymatic pathways such as xanthine oxidase, NADPH oxidase, sorbitol flux, uncoupled nitric oxide synthase (NOS), and glycolysis [11]. The NADPH oxidase pathway, which is found in mesangial cells, proximal tubular epithelia, vascular smooth muscle cells, endothelial and interstitial fibroblasts, is a major contributor to the progression of DN [27,35–40]. Subunits of NADPH oxidase were shown to be elevated in models of diabetic kidney disease and renal injury due to ROS in experimental models [41–44]. Another cytosolic origin for ROS is the uncoupling of NOS. Through different isoforms and cofactors of NOS, this uncoupling produces nitric oxide (NO) radicals [11]. It is speculated that NO plays a role in the early stages of DN through inducing hemodynamic changes [44–47]. Excess production of ROS due to hyperglycemic conditions and activated anti-nuclear factor- κ B (NF- κ B) is linked to increased inflammatory cytokine concentrations in the kidneys of diabetic rats [30,48].

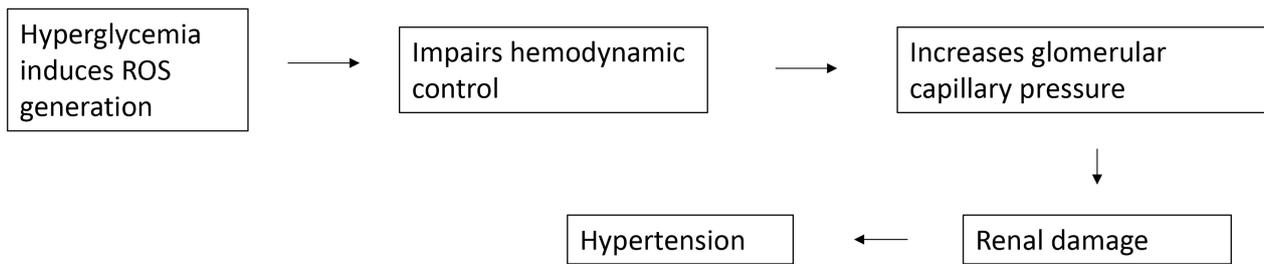


Fig. 1. Potential pathogenesis of hypertension via hyperglycemic ROS production. ROS, reactive oxygen species.

2.4 Hypertension and Oxidative Stress

Oxidative stress in the kidney and hypertension has a complex relationship (Fig. 1). Evidence has suggested that the development of hypertension can be caused by oxidative stress. This is supported by studies that show that inducing oxidative stress leads to hypertension and the depletion of extracellular superoxide dismutase (EC-SOD), an anti-oxidative enzyme, increases basal blood pressure [49–53]. Furthermore, ROS produced under hyperglycemic conditions increases vascular NO and transforming growth factor (TGF)- β 1, which is associated with vasodilation of the afferent and efferent arterioles [54–58]. Hyperglycemia has also been linked to renin-angiotensin-aldosterone system (RAAS) activation [22]. Hyperglycemia-induced RAAS activation can be inhibited via attenuation of ROS generation, suggesting that ROS generation mediates RAAS activation [59]. Dysregulated RAAS activation, in turn, can aggravate hypertension which in turn could cause renal impairment by increasing glomerular capillary pressure. Given the link between inflammation via ROS, neuro-hormonal change, and the role of the kidney in modulating blood pressure, interventions to attenuate hyperglycemia-induced oxidative stress present an opportunity to slow the progression of CKD in diabetic kidney disease.

2.5 Downstream Effects of Oxidative Stress and DN

When the overproduction of ROS occurs and there is a disproportionate amount of antioxidants, tissue damage can occur through the oxidation of protein, nucleic acids, carbohydrates, and lipids [11,12,20,60,61]. Not only does the reactivity of these species directly cause damage but ROS can act as signaling molecules, during hyperglycemia, for membrane receptor signaling to induce stress-related pathways that cause damage to cells [62]. ROS can also disrupt DNA by causing breaks in the single or double-stranded helices as well as interfering with histones [63,64]. As such, more targeted therapies specific to anti-oxidation are a potential novel treatment for DN.

3. Current Treatments of Diabetic Nephropathy and Limitations

3.1 Statin

A systematic review and meta-analysis of clinical trials evaluating five types of statins (atorvastatin, cerivastatin, lovastatin, simvastatin, pitavastatin) showed that statins significantly increased estimated glomerular filtration rate (eGFR) and reduced serum creatinine, thus improving renal function. Statin also reduces C-reactive protein (CRP) level, thus protecting the kidney from a high level of inflammation. Blood lipids are also reduced by statin, thus addressing one of the root causes of DN [65].

Previous studies have also shown statins' ability to reduce lipid deposition on the vascular wall. This is conducive to the treatment of early DN, reducing proteinuria, and delaying the progression of kidney disease [66]. Statin also improves NO activity in the vascular endothelium, improving its function and relieving contraction found in DN [67]. With respect to diabetes, statins reduce the inflammatory response and stabilize platelet function [68].

3.2 Vitamin D

Vitamin D supplementation was found to decrease albuminuria and urinary TGF- β 1 in patients with type 2 diabetes, as such, it was expected that vitamin D can protect kidneys from DN [69,70]. Follow-up studies reveal that this anti-nephropathic effect is mixed, with one study showing reduced proteinuria, prevention of kidney injury, both independent of blood pressure and blood glucose level [71]. However, this was contradicted by a study that shows no vitamin D effect on urine albumin to creatinine ratio [72]. A systematic review and meta-analysis of clinical trials examining the effects of vitamin D in DN revealed that vitamin D does improve 24-hour urine protein and inflammation indices. However, vitamin D has no effect on serum creatinine, eGFR, or glycemic control [72].

3.3 Mineralocorticoid Receptor Antagonists

Mineralocorticoid receptor antagonists (MRA) have been used to treat resistant hypertension due to high aldosterone levels [73]. Aldosterone affects sodium and potassium balance by binding to the mineralocorticoid receptor

(MR) at distal nephrons in the kidney [74]. Overactivation of MR has been found to be common in DN, driving inflammation and fibrosis in the kidney [75]. Initial MRA has been steroid-based, such as spironolactone, eplerenone, and was found to carry a risk of hyperkalemia, with contraindication for patients with serum creatinine >2 mg/dL [76,77]. More recent non-steroid MRA has a lower risk of hyperkalemia, such as finerenone, and was shown to have high selectivity toward MR, with strong anti-inflammatory and anti-fibrotic effects in the kidney, however, it is still not recommended for patients with severe CKD [78,79].

3.4 α -lipoic Acid + Valsartan

Valsartan is an angiotensin 2 receptor blocker (ARB), used to treat hypertension. Low-dose valsartan with no clinical effect on blood pressure was shown to decrease urinary albumin excretion in patients with DN. It's theorized that valsartan dilates the efferent more than afferent arterioles resulting in decreased intraglomerular pressure, protein filtration, and proteinuria [80]. In combination with angiotensin-converting enzyme (ACE) inhibitor, valsartan delays glomerular sclerosis and improves renal function in patients with type 1 DN; although, combined ARB and ACE inhibitor increases the risk for adverse drug event and are not generally used in clinical practice.

α -lipoic acid (ALA) is an antioxidant that was shown to up-regulate the expression of Nrf2-mediated antioxidant genes, peroxisome proliferator-activated receptors-regulated genes, and was used to treat diabetes [81]. In this case, ALA improved insulin sensitivity, protecting remaining β -cells by reducing their oxidative stress, reducing lipid peroxidation, and inhibiting protein glycation [81,82]. Meta-analysis of the combination of ALA and valsartan in DN showed that this combination significantly reduces urinary albumin, level of oxidative stress markers, increases antioxidant capacity, and alleviates renal function damage [83].

3.5 SGLT2 Inhibitor

Sodium-glucose cotransporter 2 (SGLT2) inhibitors, commonly known as gliflozins, are used to lower blood glucose levels by inhibiting glucose absorption at proximal tubules [84]. In the Empagliflozin Cardiovascular Outcome Event Trial in Type 2 Diabetes Mellitus Patients—Removing Excess Glucose (EMPA-REG) trial, empagliflozin reduced progression to macroalbuminuria, decreased serum creatinine, and resulted in a 39% reduction in the death rate by kidney failure [85]. The SGLT2 inhibitor also has a blood glucose-lowering effect, reducing HbA1c with its maximum effect at 6 months post-treatment, and maintaining the effect from then on up to 1 year in the clinical trial [86]. Its hypoglycemic risk is low, similar to metformin [87].

SGLT2 inhibitor improves kidney health in DN by the following actions:

(1) Restoring tubuloglomerular feedback: Increased adenosine expression, which causes vasoconstriction of the afferent arteriole, thus reducing glomerular pressure and hyperfiltration [88].

(2) Reducing tubular workload and hypoxia: Diabetic patients have enhanced proximal tubular glucose reabsorption because of their hyperglycemia, leading to increased glomerular hyperfiltration [88]. This increased reabsorption increases oxygen demand by the cells, leading to tubular hypoxia, and is a strong indication of the progression of diabetic kidney disease [88]. By decreasing glucose reabsorption, SGLT2 inhibitors lower the workload and reduce tubular hypoxia.

(3) Diuretic and natriuretic effect: SGLT2 inhibitors function as a diuretic, but with preferential fluid mobilization from the interstitial compartment rather than the intravascular compartment [89]. This helps to reduce the amount of interstitial fluid in the kidney, which may reduce cortical and medullary hypoxia.

(4) Anti-inflammatory and antifibrotic effect: SGLT2 inhibitors reduce the markers of DN: Nuclear factor- κ B (NFB), interleukin 6 (IL-6), monocyte chemoattractant protein 1 (MCP-1), and the serum uric acid level, thus reducing the risk of kidney inflammation [90,91].

3.6 Curcumin

Curcumin, found in turmeric, has strong anti-inflammatory properties that were expected to reduce inflammation-mediated kidney damage in DN. In clinical trials, it was found to have a positive impact on proteinuria, however, it has no impact on blood urea nitrogen, creatinine, eGFR, and serum albumin [92].

3.7 ACE Inhibitor/ARB (General)

Angiotensin-converting enzyme (ACE) inhibitor, or Angiotensin receptor blocker (ARB), are common hypertension medications that have also been used for the treatment of DN [93]. This is especially recommended for non-pregnant patients with both diabetes and hypertension comorbidity, as per American Diabetes Association (ADA) guideline in 2018. However, neither ACE inhibitor nor ARB is recommended for primary prevention of diabetic nephropathy, in diabetic patients with normal blood pressure, normal urinary albumin to creatinine ratio, and normal eGFR [93]. The mechanism of ACE inhibitors in diabetic nephropathy is suggested to be the lowering of glomerular intracapillary pressure, while still maintaining the flow of renal plasma. Additionally, ACE inhibitors may also prevent cellular and glomerular hypertrophy, and reduce mesangial matrix accumulation [94]. Meanwhile, ARB has been shown to prevent or delay DN independent of its action on hypertension, in patients with type 2 diabetes and microalbuminuria [95]. This data was supported by further clinical trials, except for analysis on obese or overweight patients [96].

3.8 Glycemic Control

Controlling blood sugar levels is a key requirement for living with diabetes which includes insulin treatments and healthy lifestyle habits such as diet and exercise. A strict and intense diabetic treatment plan aimed at glycemic control showed a delayed onset of diabetic retinopathy, nephropathy, and neuropathy in patients with insulin-dependent diabetes mellitus [97]. Controlling glycemic levels in adolescent patients is especially challenging as they now take responsibility for their diabetic management and still require more education, support systems, and guidance [98]. In adolescent patients, HbA1c levels were 1–2% higher in both the intensive and conventional treatments compared to adults according to the DCCT research group [97].

4. Anti-Oxidative Therapy and the Nrf2 Pathway

Identifying anti-oxidative therapies aimed at slowing and reversing the progression of CKD has become a central focus of treating DN. Largely, two approaches have been taken to restore redox balance in kidneys under diabetogenic conditions:

(1) Upregulating anti-oxidative enzymes through activation of Nrf2 pathways; (2) Reducing the production of ROS through attenuation of mitochondrial dysfunction.

The Nrf2 is a transcription factor that is centrally involved in the management of oxidative stressors throughout the body. Nrf2 is expressed in all cells at low levels under normal physiologic conditions [99]. Common sources of oxidative stress include superoxide anions, hydroxyl radicals, and hydrogen peroxide produced by mitochondria, endoplasmic reticuli (ER), and peroxisomes as a part of normal metabolic function [2,100]. The role of Nrf2 in limiting cellular damage to such ROS means that it is integral in maintaining overall cell health. As such, a dysregulation of the Nrf2 pathway results in an inability of cells to cope with oxidative stress leading to overall cellular dysfunction and disease. Studies utilizing Nrf2 deficient mice have associated Nrf2 dysregulation with a host of diseases including cancer, Alzheimer's disease, cardiovascular disease, and DN [100–102].

When oxidative stress is low, Nrf2 is bound to the E3 ubiquitin ligase subunit: Kelch-like ECH-associated protein 1 (Keap1) [102]. When complexed with Keap1, Nrf2 is targeted for ubiquitination and subsequent proteasomal degradation (Fig. 2) [103]. In this way, nuclear translocation of free Nrf2 is suppressed when ROS levels are low. When oxidative stress is high, ROS react with Keap1 releasing Nrf2 to translocate to the nucleus and bind with the antioxidant response element (ARE) promoter region (Fig. 2) [99]. Nrf2-ARE binding activates transcription of a multitude of different antioxidant, metabolic, and anti-inflammatory proteins in addition to an upregulation of Nrf2 transcription [99]. Overexpression of Nrf2 and/or loss

of Keap1 function is also associated with increased rates of autophagy [104].

Nrf2, Diabetic Nephropathy, and Anti-Oxidative Therapies

Hemodynamic and metabolic changes in individuals with diabetes result in an increase in the production of ROS which leads to alterations in renal cell structure and a decrease in kidney function [105]. In DN, the accumulation of ROS leads to an overall increase in free cytosolic Nrf2, however, Nrf2 nuclear translocation is impaired [106]. This suggests that an increase in Nrf2 production alone is not sufficient to combat the increased oxidative stress present in DN. In light of this information, Nrf2 nuclear translocation should also be a target of therapies seeking to mitigate the effects of kidney disease in individuals with DN. Mohan *et al.* [107] demonstrated the efficacy of epigallocatechin-3-gallate (ECGC) in increasing Nrf2 translocation in diabetic mice as measured by increased expression of functional HO1, γ -GCS, and NQO1. Pharmacologic ARE inducers like sulforaphane, Oltipraz, CDDO-Im, BHT increase free Nrf2 through binding of cysteine thiol groups in the Keap1 repressor [100]. Either ECGC, ARE inducers, or a combination thereof could hold promise as potential therapies for DN. Evidence also shows that Nrf2 activators have been shown to slow the progression of DN [105,108–114]. For example, dimethyl fumarate (DMF) is an Nrf2 activator with anti-oxidative, anti-inflammatory, and immunomodulating effects that are currently used as a treatment for psoriasis and relapsing-remitting multiple sclerosis [115]. In a study of 30 diabetic mice, different doses of DMF, low, medium, and high were given for three months to assess DN, lipid profile, renal hypertrophy, oxidative stress in the kidney, and renal histopathological changes [116]. It was found that high dose DMF (80 mg/kg) was able to attenuate DN significantly [117].

While increasing anti-oxidative enzyme expression is an important aspect of treating DN, reducing the production of ROS and inflammation is also important in the long-term management of DN. Diabetogenic conditions result in abnormal metabolism of glucose and fatty acids by the mitochondria, resulting in dysfunction and activation of NADPH oxidase, one of the major contributors of ROS production [118].

Several other macromolecules and pathways have been implicated in ROS production including advanced glycation end products (AGE), defects in polyol pathway, and uncoupled nitric oxide synthase (NOS). In this review, we focus on mitochondrial dysfunction and therapeutic agents targeted at restoring mitochondrial homeostasis as an important point of intervention to suppress ROS production [25]. Early diabetic kidney disease has been associated with mitochondrial fragmentation, fission, impaired mitophagy, decreased mitochondrial membrane potential, and ROS production [119]. Several agents such as Tilapia Skin Peptides, berberine, Sirte 6, and progranulin have been

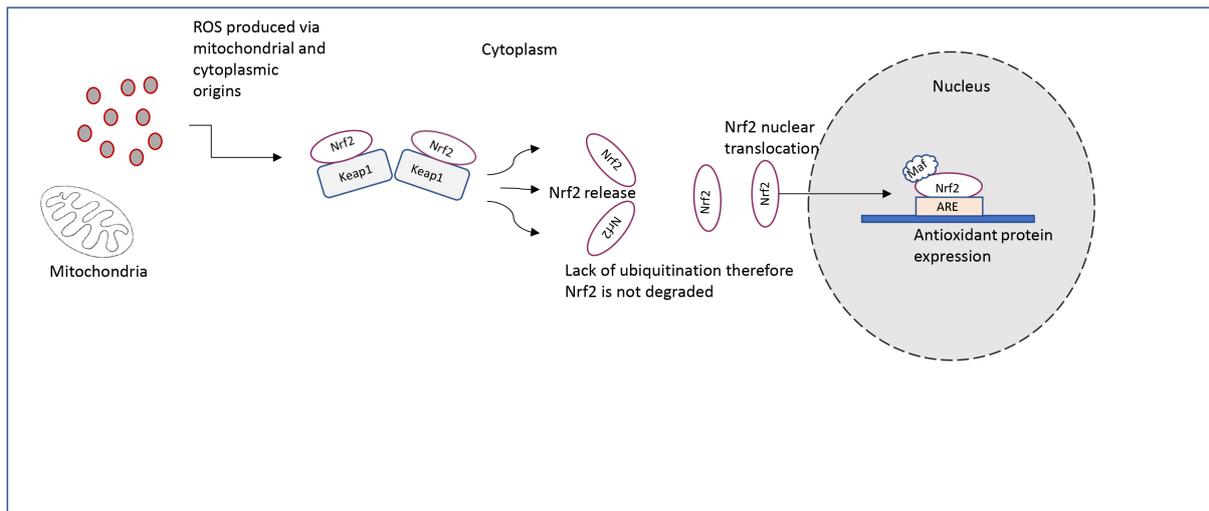


Fig. 2. The Nrf2 and Keap1 pathway under oxidative stress conditions. Under non-stressed conditions, Nrf2 is ubiquitinated and degraded in the proteasome. Under stressed conditions, ubiquitination is downregulated and Nrf2 is free to translocate to the nucleus.

shown to impede the progression of DN by restoring mitochondrial morphology and function [120–125]. In particular, pyruvate kinase M2, a glucose metabolic enzyme, was found to be elevated in kidneys of diabetic patients who did not develop DN but remained low in patients with progressed to DN [120]. Pyruvate kinase muscle isozyme 2 (PKM2) activation by TEPP-46 reversed diabetes-induced defects in mitochondrial function and prevented the development of glomerular pathology evidenced by a mouse model of DN [120]. Unfortunately, these agents are yet to be studied in humans and thus the safety and efficacy are unknown. Nrf2 activators, in addition to elevating the expression of antioxidants, have also been shown to exhibit their therapeutic effects by restoring mitochondrial homeostasis. Human proximal tubular cells, human kidney-2 (HK-2) cells, cultured under hypoxic ambiance to emulate the hypoxic state under diabetic conditions resulted in increased mitochondrial fragmentation, ROS production, mitochondrial membrane potential loss, and apoptosis; these results were reversed with cotreatment with an HO-1 agonist [125]. Furthermore, MitoQ and CoQ10, mitochondrial antioxidants restored mitophagy and mitochondrial quality control *in vivo* in streptozotocin-induced DN [107,117]. Thus, further advancements and identification of Nrf2 activators that also restores mitochondrial homeostasis is a promising approach in the treatment and prevention of DN.

5. Conclusions

DN and oxidative stress are intertwined in CKD progression. The overproduction of ROS causes vascular damage and over time impairs the kidney. Hyperglycemia further aggravates oxidative stress by inducing more production of ROS. Current treatments have long been known to delay the progression of diabetic kidney disease; how-

ever, the majority of patients with DN progress to ESRD. Therefore, targeted therapies on the antioxidation via the Nrf2 pathway can be a promising option. Ultimately, a multimodal therapeutic approach involving traditional and newer anti-oxidative therapies applicable in clinical practice should be further explored to mitigate CKD progression in patients with DN.

Author Contributions

HI (Author F) conceived the idea of writing a review on DN and anti-oxidative stress. LFH (Author A) developed the idea into a draft outline and wrote the abstract, introduction, DN and Oxidative stress section, conclusion, and made the tables and figures. NE (Author B) wrote about current anti-oxidative treatments, assisted in the organization of the paper, and provided help and advice in the development of the paper. DW (Author C) wrote about the Nrf2 pathway, offered advice, and assisted in the development of the paper. MA (Author D) wrote the current treatment and limitations section and assisted in the development of the paper. LFH, NE, DW, MA and ET (Author A, B, C, D, and E) wrote the manuscript. All authors contributed to editorial changes in the manuscript. All authors read and approved the final manuscript.

Ethics Approval and Consent to Participate

Not applicable.

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Conflict of Interest

The authors declare no conflict of interest.

References

- [1] Rabkin R. Diabetic nephropathy. *Clinical Cornerstone*. 2003; 5: 1–11.
- [2] Cao Z, Cooper ME. Pathogenesis of diabetic nephropathy. *Journal of Diabetes Investigation*. 2011; 2: 243–247.
- [3] Ritz E. Limitations and future treatment options in type 2 diabetes with renal impairment. *Diabetes Care*. 2011; 34: S330–S334.
- [4] Tavafi M. Diabetic nephropathy and antioxidants. *Journal of Nephropathology*. 2013; 2: 20–27.
- [5] Bossy-Wetzel E, Schwarzenbacher R, Lipton SA. Molecular pathways to neurodegeneration. *Nature Medicine*. 2004; 10: S2–S9.
- [6] Balaban RS, Nemoto S, Finkel T. Mitochondria, Oxidants, and Aging. *Cell*. 2005; 120: 483–495.
- [7] Ma Q. Role of nrf2 in oxidative stress and toxicity. *Annual Review of Pharmacology and Toxicology*. 2013; 53: 401–426.
- [8] Welt K, Weiss J, Martin R, Hermsdorf T, Drews S, Fitzl G. Ginkgo biloba extract protects rat kidney from diabetic and hypoxic damage. *Phytomedicine*. 2007; 14: 196–203.
- [9] Al-Qattan KK, Thomson M, Jayasree D, Ali M. Garlic Attenuates Plasma and Kidney ACE-I and AngII Modulations in Early Streptozotocin-Induced Diabetic Rats: Renal Clearance and Blood Pressure Implications. *Evidence-Based Complementary and Alternative Medicine*. 2016; 2016: 1–11.
- [10] Cooper ME. Pathogenesis, prevention, and treatment of diabetic nephropathy. *The Lancet*. 1998; 352: 213–219.
- [11] Forbes JM, Coughlan MT, Cooper ME. Oxidative stress as a major culprit in kidney disease in diabetes. *Diabetes*. 2008; 57: 1446–1454.
- [12] Brownlee M. The pathobiology of diabetic complications: a unifying mechanism. *Diabetes*. 2005; 54: 1615–1625.
- [13] Gnudi L, Thomas SM, Viberti G. Mechanical Forces in Diabetic Kidney Disease: a Trigger for Impaired Glucose Metabolism. *Journal of the American Society of Nephrology*. 2007; 18: 2226–2232.
- [14] Gorelik Y, Bloch-Isenberg N, Hashoul S, Heyman SN, Khamaisi M. Hyperglycemia on Admission Predicts Acute Kidney Failure and Renal Functional Recovery among Inpatients. *Journal of Clinical Medicine*. 2021; 11: 54.
- [15] Hostetter TH, Troy JL, Brenner BM. Glomerular hemodynamics in experimental diabetes mellitus. *Kidney International*. 1981; 19: 410–415.
- [16] Zatz R, Dunn BR, Meyer TW, Anderson S, Rennke HG, Brenner BM. Prevention of diabetic glomerulopathy by pharmacological amelioration of glomerular capillary hypertension. *Journal of Clinical Investigation*. 1986; 77: 1925–1930.
- [17] Keane WF, Kasiske BL, O'Donnell MP, Kim Y. Hypertension, Hyperlipidemia, and Renal Damage. *American Journal of Kidney Diseases*. 1993; 21: 43–50.
- [18] Tangvarasittichai S. Oxidative stress, insulin resistance, dyslipidemia and type 2 diabetes mellitus. *World Journal of Diabetes*. 2015; 6: 456–480.
- [19] Li R, Jia Z, Trush MA. Defining ROS in Biology and Medicine. *The Reactive Oxygen Species (Apex)*. 2016; 1: 9–21.
- [20] Rosen P, Nawroth PP, King G, Moller W, Tritschler HJ, Packer L. The role of oxidative stress in the onset and progression of diabetes and its complications: a summary of a Congress Series sponsored by UNESCO-MCBN, the American Diabetes Association and the German Diabetes Society. *Diabetes/Metabolism Research and Reviews*. 2001; 17: 189–212.
- [21] Gnudi L. Cellular and molecular mechanisms of diabetic glomerulopathy. *Nephrology Dialysis Transplantation*. 2012; 27: 2642–2649.
- [22] Chen C, Juan S, Chou H. Hyperglycemia activates the renin-angiotensin system and induces epithelial-mesenchymal transition in streptozotocin-induced diabetic kidneys. *Journal of the Renin-Angiotensin-Aldosterone System*. 2018; 19: 147032031880300.
- [23] Ceriello A, Morocutti A, Mercuri F, Quagliaro L, Moro M, Damante G, *et al.* Defective intracellular antioxidant enzyme production in type 1 diabetic patients with nephropathy. *Diabetes*. 2000; 49: 2170–2177.
- [24] Baynes JW, Thorpe SR. Role of oxidative stress in diabetic complications: a new perspective on an old paradigm. *Diabetes*. 1999; 48: 1–9.
- [25] Kashiwara N, Haruna Y, K. Kondeti V, S. Kanwar Y. Oxidative Stress in Diabetic Nephropathy. *Current Medicinal Chemistry*. 2010; 17: 4256–4269.
- [26] Vasavada N, Agarwal R. Role of oxidative stress in diabetic nephropathy. *Advances in Chronic Kidney Disease*. 2005; 12: 146–154.
- [27] Gill PS, Wilcox CS. NADPH Oxidases in the Kidney. *Antioxidants & Redox Signaling*. 2006; 8: 1597–1607.
- [28] Riemer J, Bulleid N, Herrmann JM. Disulfide Formation in the ER and Mitochondria: Two Solutions to a Common Process. *Science*. 2009; 324: 1284–1287.
- [29] Addabbo F, Montagnani M, Goligorsky MS. Mitochondria and Reactive Oxygen Species. *Hypertension*. 2009; 53: 885–892.
- [30] Kowaltowski AJ, de Souza-Pinto NC, Castilho RF, Vercesi AE. Mitochondria and reactive oxygen species. *Free Radical Biology and Medicine*. 2009; 47: 333–343.
- [31] Kabat A, Pönicke K, Salameh A, Friedrich-Wilhelm M, Dhein S. Effect of a beta 2-adrenoceptor stimulation on hyperglycemia-induced endothelial dysfunction. *The Journal of Pharmacology and Experimental Therapeutics*. 2004; 308: 564–573.
- [32] Nishikawa T, Edelstein D, Du XL, Yamagishi S, Matsumura T, Kaneda Y, *et al.* Normalizing mitochondrial superoxide production blocks three pathways of hyperglycaemic damage. *Nature*. 2000; 404: 787–790.
- [33] Yamagishi S, Inagaki Y, Okamoto T, Amano S, Koga K, Takeuchi M, *et al.* Advanced Glycation End Product-induced Apoptosis and Overexpression of Vascular Endothelial Growth Factor and Monocyte Chemoattractant Protein-1 in Human-cultured Mesangial Cells. *Journal of Biological Chemistry*. 2002; 277: 20309–20315.
- [34] Nowotny K, Jung T, Höhn A, Weber D, Grune T. Advanced glycation end products and oxidative stress in type 2 diabetes mellitus. *Biomolecules*. 2015; 5: 194–222.
- [35] Li J, Shah AM. ROS Generation by Nonphagocytic NADPH Oxidase: Potential Relevance in Diabetic Nephropathy. *Journal of the American Society of Nephrology*. 2003; 14: S221–S226.
- [36] Zalba G, José GS, Moreno MU, Fortuño MA, Fortuño A, Beaumont FJ, *et al.* Oxidative Stress in Arterial Hypertension: role of NAD(P)H oxidase. *Hypertension*. 2001; 38: 1395–1399.
- [37] Yang S, Madyastha P, Bingel S, Ries W, Key L. A New Superoxide-generating Oxidase in Murine Osteoclasts. *Journal of Biological Chemistry*. 2001; 276: 5452–5458.
- [38] Jones SA, Hancock JT, Jones OT, Neubauer A, Topley N. The expression of NADPH oxidase components in human glomerular mesangial cells: detection of protein and mRNA for p47phox, p67phox, and p22phox. *Journal of the American Society of Nephrology*. 1995; 5: 1483–1491.
- [39] Shiose A, Kuroda J, Tsuruya K, Hirai M, Hirakata H, Naito S, *et*

- al.* A Novel Superoxide-producing NAD(P)H Oxidase in Kidney. *Journal of Biological Chemistry*. 2001; 276: 1417–1423.
- [40] Radeke HH, Cross AR, Hancock JT, Jones OT, Nakamura M, Kaefer V, *et al.* Functional expression of NADPH oxidase components (alpha- and beta-subunits of cytochrome b558 and 45-kDa flavoprotein) by intrinsic human glomerular mesangial cells. *Journal of Biological Chemistry*. 1991; 266: 21025–21029.
- [41] Asaba K, Tojo A, Onozato ML, Goto A, Quinn MT, Fujita T, *et al.* Effects of NADPH oxidase inhibitor in diabetic nephropathy. *Kidney International*. 2005; 67: 1890–1898.
- [42] Nam SM, Lee MY, Koh JH, Park JH, Shin JY, Shin YG, *et al.* Effects of NADPH oxidase inhibitor on diabetic nephropathy in OLETF rats: the role of reducing oxidative stress in its protective property. *Diabetes Research and Clinical Practice*. 2009; 83: 176–182.
- [43] Thallas-Bonke V, Thorpe SR, Coughlan MT, Fukami K, Yap FY, Sourris KC, *et al.* Inhibition of NADPH oxidase prevents advanced glycation end product-mediated damage in diabetic nephropathy through a protein kinase C-alpha-dependent pathway. *Diabetes*. 2008; 57: 460–469.
- [44] Han HJ, Lee YJ, Park SH, Lee JH, Taub M. High glucose-induced oxidative stress inhibits Na⁺/glucose cotransporter activity in renal proximal tubule cells. *The American Journal of Physiology-Renal Physiology*. 2005; 288: F988–F996.
- [45] Choi KC, Lee SC, Kim SW, Kim NH, Lee JU, Kang YJ. Role of Nitric Oxide in the Pathogenesis of Diabetic Nephropathy in Streptozotocin - Induced Diabetic Rats. *The Korean Journal of Internal Medicine*. 1999; 14: 32–41.
- [46] Satoh M, Fujimoto S, Haruna Y, Arakawa S, Horike H, Komai N, *et al.* NAD(P)H oxidase and uncoupled nitric oxide synthase are major sources of glomerular superoxide in rats with experimental diabetic nephropathy. *American Journal of Physiology-Renal Physiology*. 2005; 288: F1144–F1152.
- [47] Komers R, Allen TJ, Cooper ME. Role of endothelium-derived nitric oxide in the pathogenesis of the renal hemodynamic changes of experimental diabetes. *Diabetes*. 1994; 43: 1190–1197.
- [48] Bhattacharya S, Manna P, Gachhui R, Sil PC. D-saccharic acid 1,4-lactone protects diabetic rat kidney by ameliorating hyperglycemia-mediated oxidative stress and renal inflammatory cytokines via NF-kappaB and PKC signaling. *Toxicology and Applied Pharmacology*. 2013; 267: 16–29.
- [49] Wilcox CS. Oxidative stress and nitric oxide deficiency in the kidney: a critical link to hypertension? *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*. 2005; 289: R913–R935.
- [50] Ni Z, Hou S, Barton CH, Vaziri ND. Lead exposure raises superoxide and hydrogen peroxide in human endothelial and vascular smooth muscle cells. *Kidney International*. 2004; 66: 2329–2336.
- [51] Ding Y, Gonick HC, Vaziri ND. Lead promotes hydroxyl radical generation and lipid peroxidation in cultured aortic endothelial cells. *American Journal of Hypertension*. 2000; 13: 552–555.
- [52] Chabrashvili T, Tojo A, Onozato ML, Kitiyakara C, Quinn MT, Fujita T, *et al.* Expression and Cellular Localization of Classic NADPH Oxidase Subunits in the Spontaneously Hypertensive Rat Kidney. *Hypertension*. 2002; 39: 269–274.
- [53] Chu Y, Iida S, Lund DD, Weiss RM, DiBona GF, Watanabe Y, *et al.* Gene Transfer of Extracellular Superoxide Dismutase Reduces Arterial Pressure in Spontaneously Hypertensive Rats: role of heparin-binding domain. *Circulation Research*. 2003; 92: 461–468.
- [54] Sagoo MK, Gnudi L. Diabetic nephropathy: is there a role for oxidative stress? *Free Radical Biology and Medicine*. 2018; 116: 50–63.
- [55] De Vriese AS, Stoenoiu MS, Elger M, Devuyst O, Vanholder R, Kriz W, *et al.* Diabetes-induced microvascular dysfunction in the hydronephrotic kidney: Role of nitric oxide. *Kidney International*. 2001; 60: 202–210.
- [56] Kagami S. Involvement of glomerular renin-angiotensin system (RAS) activation in the development and progression of glomerular injury. *Clinical and Experimental Nephrology*. 2012; 16: 214–220.
- [57] RAIJ L. The pathophysiologic basis for blocking the renin-angiotensin system in hypertensive patients with renal disease. *American Journal of Hypertension*. 2005; 18: 95–99.
- [58] Sharma K, Cook A, Smith M, Valancius C, Inscho EW. TGF-beta impairs renal autoregulation via generation of ROS. *American Journal of Physiology-Renal Physiology*. 2005; 288: F1069–F1077.
- [59] Xue H, Yuan P, Ni J, Li C, Shao D, Liu J, *et al.* H(2)S inhibits hyperglycemia-induced intrarenal renin-angiotensin system activation via attenuation of reactive oxygen species generation. *PLoS ONE*. 2013; 8: e74366.
- [60] Papaharalambus CA, Griendling KK. Basic Mechanisms of Oxidative Stress and Reactive Oxygen Species in Cardiovascular Injury. *Trends in Cardiovascular Medicine*. 2007; 17: 48–54.
- [61] Kenneth Maiese, Zhao Zhong Chong, Yan Chen Shang. Mechanistic Insights into Diabetes Mellitus and Oxidative Stress. *Current Medicinal Chemistry*. 2007; 14: 1729–1738.
- [62] Ha H, Lee HB. Reactive oxygen species as glucose signaling molecules in mesangial cells cultured under high glucose. *Kidney International*. 2000; 58: S19–S25.
- [63] Esposito LA, Melov S, Panov A, Cottrell BA, Wallace DC. Mitochondrial disease in mouse results in increased oxidative stress. *Proceedings of the National Academy of Sciences*. 1999; 96: 4820–4825.
- [64] Lieber MR, Karanjawala ZE. Ageing, repetitive genomes and DNA damage. *Nature Reviews Molecular Cell Biology*. 2004; 5: 69–75.
- [65] Lv J, Ren C, Hu Q. Effect of statins on the treatment of early diabetic nephropathy: a systematic review and meta-analysis of nine randomized controlled trials. *Annals of Palliative Medicine*. 2021; 10: 11548–11557.
- [66] Zhang L, Zhang S, Yu Y, Jiang H, Ge J. Efficacy and safety of rosuvastatin vs. atorvastatin in lowering LDL cholesterol: A meta-analysis of trials with East Asian populations. *Herz*. 2020; 45: 594–602.
- [67] Takazakura A, Sakurai M, Bando Y, Misu H, Takeshita Y, Kita Y, *et al.* Renoprotective effects of atorvastatin compared with pravastatin on progression of early diabetic nephropathy. *Journal of Diabetes Investigation*. 2015; 6: 346–353.
- [68] Bonaterra GA, Bender K, Wilhelm B, Schwarzbach H, Metz S, Kelber O, *et al.* Effect of cholesterol re-supplementation and atorvastatin on plaque composition in the thoracic aorta of New Zealand white rabbits. *BMC Cardiovascular Disorders*. 2020; 20: 420.
- [69] Kim MJ, Frankel AH, Donaldson M, Darch SJ, Pusey CD, Hill PD, *et al.* Oral cholecalciferol decreases albuminuria and urinary TGF-beta1 in patients with type 2 diabetic nephropathy on established renin-angiotensin-aldosterone system inhibition. *Kidney International*. 2011; 80: 851–860.
- [70] Zhao J, Dong J, Wang H, Shang H, Zhang D, Liao L. Efficacy and safety of vitamin D3 in patients with diabetic nephropathy: a meta-analysis of randomized controlled trials. *Chinese Medical Journal*. 2014; 127: 2837–2843.
- [71] Derakhshanian H, Shab-Bidar S, Speakman JR, Nadimi H, Djafarian K. Vitamin D and diabetic nephropathy: a systematic review and meta-analysis. *Nutrition*. 2015; 31: 1189–1194.
- [72] Wang Y, Yang S, Zhou Q, Zhang H, Yi B. Effects of Vitamin D Supplementation on Renal Function, Inflammation and

Glycemic Control in Patients with Diabetic Nephropathy: a Systematic Review and Meta-Analysis. *Kidney and Blood Pressure Research*. 2019; 44: 72–87.

- [73] Wisniewski KE, Dalton AJ, McLachlan DRC, Wen GY, Wisniewski HM. Alzheimer's disease in down's syndrome: Clinicopathologic studies. *Neurology*. 1985; 35: 957–957.
- [74] Good DW. Nongenomic Actions of Aldosterone on the Renal Tubule. *Hypertension*. 2007; 49: 728–739.
- [75] Goenka L, Padmanaban R, George M. The Ascent of Mineralocorticoid Receptor Antagonists in Diabetic Nephropathy. *Current Clinical Pharmacology*. 2019; 14: 78–83.
- [76] Bolognani D, Palmer SC, Navaneethan SD, Strippoli GF. Aldosterone antagonists for preventing the progression of chronic kidney disease. *Cochrane Database of Systematic Reviews*. 2014; 2014: CD007004.
- [77] Currie G, Taylor AHM, Fujita T, Ohtsu H, Lindhardt M, Rossing P, *et al*. Effect of mineralocorticoid receptor antagonists on proteinuria and progression of chronic kidney disease: a systematic review and meta-analysis. *BMC Nephrology*. 2016; 17: 127.
- [78] Kawanami D, Takashi Y, Muta Y, Oda N, Nagata D, Takahashi H, *et al*. Mineralocorticoid Receptor Antagonists in Diabetic Kidney Disease. *Frontiers in Pharmacology*. 2021; 12: 754239.
- [79] Baran W, Krzeminska J, Szlagor M, Wronka M, Mlynarska E, Franczyk B, *et al*. Mineralocorticoid Receptor Antagonists-Use in Chronic Kidney Disease. *International Journal of Molecular Sciences*. 2021; 22: 9995.
- [80] Suzuki K, Souda S, Ikarashi T, Kaneko S, Nakagawa O, Aizawa Y. Renoprotective effects of low-dose valsartan in type 2 diabetic patients with diabetic nephropathy. *Diabetes Research and Clinical Practice*. 2002; 57: 179–183.
- [81] Golbidi S, Badran M, Laher I. Diabetes and alpha lipoic Acid. *Frontiers in Pharmacology*. 2011; 2: 69.
- [82] Noori N, Tabibi H, Hosseinpanah F, Hedayati M, Nafar M. Effects of Combined Lipoic Acid and Pyridoxine on Albuminuria, Advanced Glycation End-Products, and Blood Pressure in Diabetic Nephropathy. *International Journal for Vitamin and Nutrition Research*. 2013; 83: 77–85.
- [83] Sun F, Jiang D, Cai J. Effects of valsartan combined with alpha-lipoic acid on renal function in patients with diabetic nephropathy: a systematic review and meta-analysis. *BMC Endocrine Disorders*. 2021; 21: 178.
- [84] Nespoux J, Vallon V. SGLT2 inhibition and kidney protection. *Clinical Science*. 2018; 132: 1329–1339.
- [85] Sharaf El Din UAA, Salem MM, Abdulazim DO. Sodium-glucose cotransporter 2 inhibitors as the first universal treatment of chronic kidney disease. *Nefrologia*. 2021. (in press)
- [86] Hsia DS, Grove O, Cefalu WT. An update on sodium-glucose co-transporter-2 inhibitors for the treatment of diabetes mellitus. *Current Opinion in Endocrinology & Diabetes and Obesity*. 2017; 24: 73–79.
- [87] Monami M, Nardini C, Mannucci E. Efficacy and safety of sodium glucose co-transport-2 inhibitors in type 2 diabetes: a meta-analysis of randomized clinical trials. *Diabetes, Obesity and Metabolism*. 2014; 16: 457–466.
- [88] Heerspink HJL, Perkins BA, Fitchett DH, Husain M, Cherney DZI. Sodium Glucose Cotransporter 2 Inhibitors in the Treatment of Diabetes Mellitus: Cardiovascular and Kidney Effects, Potential Mechanisms, and Clinical Applications. *Circulation*. 2016; 134: 752–772.
- [89] Hallow KM, Helmlinger G, Greasley PJ, McMurray JJV, Boulton DW. Why do SGLT2 inhibitors reduce heart failure hospitalization? a differential volume regulation hypothesis. *Diabetes, Obesity and Metabolism*. 2018; 20: 479–487.
- [90] Heerspink HJL, Perco P, Mulder S, Leierer J, Hansen MK, Heinzel A, *et al*. Canagliflozin reduces inflammation and fibrosis biomarkers: a potential mechanism of action for beneficial effects of SGLT2 inhibitors in diabetic kidney disease. *Diabetologia*. 2019; 62: 1154–1166.
- [91] Zhao Y, Xu L, Tian D, Xia P, Zheng H, Wang L, *et al*. Effects of sodium-glucose co-transporter 2 (SGLT2) inhibitors on serum uric acid level: A meta-analysis of randomized controlled trials. *Diabetes, Obesity and Metabolism*. 2018; 20: 458–462.
- [92] Bagherniya M, Soleimani D, Rouhani MH, Askari G, Sathya-palan T, Sahebkar A. The Use of Curcumin for the Treatment of Renal Disorders: a Systematic Review of Randomized Controlled Trials. *Studies on Biomarkers and New Targets in Aging Research in Iran*. 2021; 11: 327–343.
- [93] American Diabetes Association. Standards of Medical Care in Diabetes-2018 Abridged for Primary Care Providers. *Clinical Diabetes*. 2018; 36: 14–37.
- [94] Lewis EJ, Hunsicker LG, Bain RP, Rohde RD. The Effect of Angiotensin-Converting-Enzyme Inhibition on Diabetic Nephropathy. *New England Journal of Medicine*. 1993; 329: 1456–1462.
- [95] Parving H, Andersen S, Jacobsen P, Christensen PK, Rossing K, Hovind P, *et al*. Angiotensin receptor blockers in diabetic nephropathy: renal and cardiovascular end points. *Seminars in Nephrology*. 2004; 24: 147–157.
- [96] Sharma AM, Weir MR. The Role of Angiotensin Receptor Blockers in Diabetic Nephropathy. *Postgraduate Medicine*. 2011; 123: 109–121.
- [97] Diabetes Control and Complications Trial Research Group, Nathan DM, Genuth S, Lachin J, Cleary P, Crofford O, *et al*. The effect of intensive treatment of diabetes on the development and progression of long-term complications in insulin-dependent diabetes mellitus. *New England Journal of Medicine*. 1993; 329: 977–986.
- [98] Cravedi P, Ruggenenti P, Remuzzi A, Remuzzi G. Current Status of Islet Transplantation. *Regenerative Medicine Applications in Organ Transplantation*. 2014; 343: 583–598.
- [99] Baird L, Yamamoto M. The Molecular Mechanisms Regulating the KEAP1-NRF2 Pathway. *Molecular and Cellular Biology*. 2020; 40
- [100] Center for Disease Control and Prevention. National Diabetes Statistics Report 2020. Available at: <https://www.scirp.org/reference/referencespapers.aspx?referenceid=3056532> (Accessed: 1 February 2022).
- [101] Dodson M, de la Vega MR, Cholanians AB, Schmidlin CJ, Chapman E, Zhang DD. Modulating NRF2 in Disease: Timing is everything. *Annual Review of Pharmacology and Toxicology*. 2019; 59: 555–575.
- [102] He F, Ru X, Wen T. NRF2, a Transcription Factor for Stress Response and Beyond. *International Journal of Molecular Sciences*. 2020; 21: 4777.
- [103] Kansanen E, Kuosmanen SM, Leinonen H, Levenon A. The Keap1-Nrf2 pathway: Mechanisms of activation and dysregulation in cancer. *Redox Biology*. 2013; 1: 45–49.
- [104] Komatsu M, Kurokawa H, Waguri S, Taguchi K, Kobayashi A, Ichimura Y, *et al*. The selective autophagy substrate p62 activates the stress responsive transcription factor Nrf2 through inactivation of Keap1. *Nature Cell Biology*. 2010; 12: 213–223.
- [105] Mohan T, Narasimhan KKS, Ravi DB, Velusamy P, Chandrasekar N, Chakrapani LN, *et al*. Role of Nrf2 dysfunction in the pathogenesis of diabetic nephropathy: Therapeutic prospect of epigallocatechin-3-gallate. *Free Radical Biology and Medicine*. 2020; 160: 227–238.
- [106] Sharma P, Jha AB, Dubey RS, Pessarakli M. Reactive Oxygen Species, Oxidative Damage, and Antioxidative Defense Mechanism in Plants under Stressful Conditions. *Journal of Botany*. 2012; 2012: 1–26.
- [107] Sun J, Zhu H, Wang X, Gao Q, Li Z, Huang H. CoQ10 ameliorates mitochondrial dysfunction in diabetic nephropathy through

- mitophagy. *Journal of Endocrinology*. 2019; 240: 445–465.
- [108] Li S, Zheng L, Zhang J, Liu X, Wu Z. Inhibition of ferroptosis by up-regulating Nrf2 delayed the progression of diabetic nephropathy. *Free Radical Biology and Medicine*. 2021; 162: 435–449.
- [109] Xing L, Guo H, Meng S, Zhu B, Fang J, Huang J, *et al*. Klotho ameliorates diabetic nephropathy by activating Nrf2 signaling pathway in podocytes. *Biochemical and Biophysical Research Communications*. 2021; 534: 450–456.
- [110] Chen Y, Kong L, Tang Z, Zhang Y, Liu Y, Wang T, *et al*. Hesperetin ameliorates diabetic nephropathy in rats by activating Nrf2/are/glyoxalase 1 pathway. *Biomedicine & Pharmacotherapy*. 2019; 111: 1166–1175.
- [111] Sun W, Liu X, Zhang H, Song Y, Li T, Liu X, *et al*. Epigallocatechin gallate upregulates NRF2 to prevent diabetic nephropathy via disabling KEAP1. *Free Radical Biology and Medicine*. 2017; 108: 840–857.
- [112] Dong W, Jia Y, Liu X, Zhang H, Li T, Huang W, *et al*. Sodium butyrate activates NRF2 to ameliorate diabetic nephropathy possibly via inhibition of HDAC. *Journal of Endocrinology*. 2017; 232: 71–83.
- [113] Li Z, Guo H, Li J, Ma T, Zhou S, Zhang Z, *et al*. Sulforaphane prevents type 2 diabetes-induced nephropathy via AMPK-mediated activation of lipid metabolic pathways and Nrf2 antioxidative function. *Clinical Science*. 2020; 134: 2469–2487.
- [114] Su L, Cao P, Wang H. Tetrandrine mediates renal function and redox homeostasis in a streptozotocin-induced diabetic nephropathy rat model through Nrf2/HO-1 reactivation. *Annals of Translational Medicine*. 2020; 8: 990–990.
- [115] Kourakis S, Timpani CA, de Haan JB, Gueven N, Fischer D, Rybalka E. Dimethyl Fumarate and Its Esters: A Drug with Broad Clinical Utility? *Pharmaceuticals*. 2020; 13: 306.
- [116] Lone A, Behl T, Kumar A, Makkar R, Nijhawan P, Redhu S, *et al*. Renoprotective potential of dimethyl fumarate in streptozotocin induced diabetic nephropathy in Wistar rats. *Obesity Medicine*. 2020; 18: 100237.
- [117] Xiao L, Xu X, Zhang F, Wang M, Xu Y, Tang D, *et al*. The mitochondria-targeted antioxidant MitoQ ameliorated tubular injury mediated by mitophagy in diabetic kidney disease via Nrf2/PINK1. *Redox Biology*. 2017; 11: 297–311.
- [118] Mima A. Inflammation and Oxidative Stress in Diabetic Nephropathy: New Insights on its Inhibition as New Therapeutic Targets. *Journal of Diabetes Research*. 2013; 2013: 248563.
- [119] Wei PZ, Szeto CC. Mitochondrial dysfunction in diabetic kidney disease. *Clinica Chimica Acta*. 2019; 496: 108–116.
- [120] Qi W, Keenan HA, Li Q, Ishikado A, Kannt A, Sadowski T, *et al*. Pyruvate kinase M2 activation may protect against the progression of diabetic glomerular pathology and mitochondrial dysfunction. *Nature Medicine*. 2017; 23: 753–762.
- [121] Qin X, Zhao Y, Gong J, Huang W, Su H, Yuan F, *et al*. Berberine Protects Glomerular Podocytes via Inhibiting Drp1-Mediated Mitochondrial Fission and Dysfunction. *Theranostics*. 2019; 9: 1698–1713.
- [122] Jin L, Zheng D, Yang G, Li W, Yang H, Jiang Q, *et al*. Tilapia Skin Peptides Ameliorate Diabetic Nephropathy in STZ-Induced Diabetic Rats and HG-Induced GMCs by Improving Mitochondrial Dysfunction. *Marine Drugs*. 2020; 18: 363.
- [123] Zhou D, Zhou M, Wang Z, Fu Y, Jia M, Wang X, *et al*. PGRN acts as a novel regulator of mitochondrial homeostasis by facilitating mitophagy and mitochondrial biogenesis to prevent podocyte injury in diabetic nephropathy. *Cell Death & Disease*. 2019; 10: 524.
- [124] Fan Y, Yang Q, Yang Y, Gao Z, Ma Y, Zhang L, *et al*. Sirt6 Suppresses High Glucose-Induced Mitochondrial Dysfunction and Apoptosis in Podocytes through AMPK Activation. *International Journal of Biological Sciences*. 2019; 15: 701–713.
- [125] Jiang N, Zhao H, Han Y, Li L, Xiong S, Zeng L, *et al*. HIF-1 α ameliorates tubular injury in diabetic nephropathy via HO-1-mediated control of mitochondrial dynamics. *Cell Proliferation*. 2020; 53: e12909.